

## CM201 - October 4, 2022

Item # CM201 was discontinued on October 4, 2022. For informational purposes, this is a copy of the website content at that time and is valid only for the stated product.

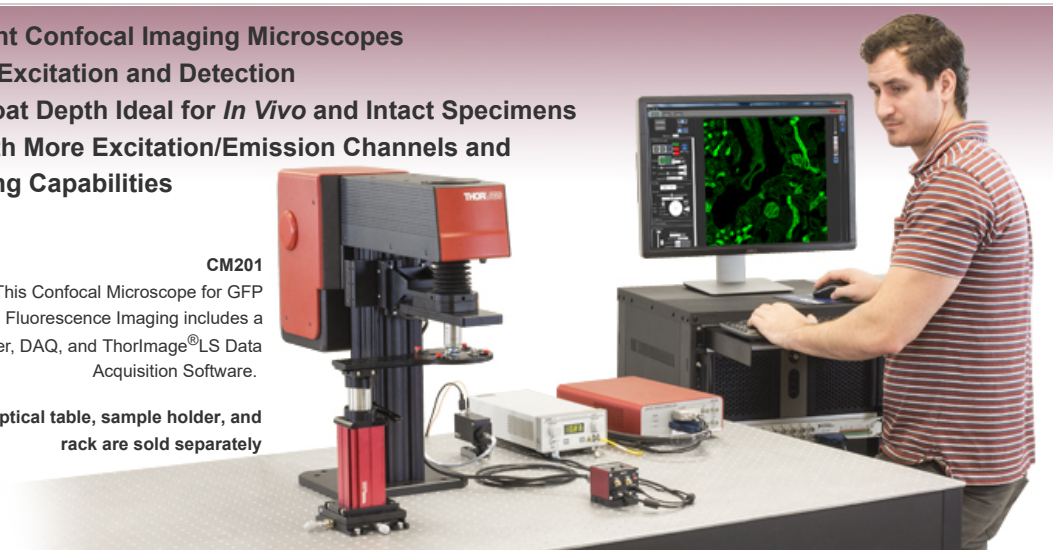
### UPGRADEABLE SINGLE-CHANNEL, CERNA®-BASED CONFOCAL SYSTEMS

- ▶ Complete Upright Confocal Imaging Microscopes
- ▶ Single-Channel Excitation and Detection
- ▶ Large 7.74" Throat Depth Ideal for *In Vivo* and Intact Specimens
- ▶ Upgradeable with More Excitation/Emission Channels and Widefield Imaging Capabilities

#### CM201

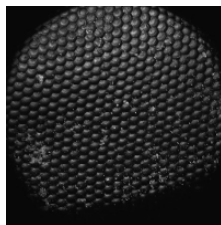
This Confocal Microscope for GFP Fluorescence Imaging includes a computer, DAQ, and ThorImage®LS Data Acquisition Software.

The optical table, sample holder, and rack are sold separately

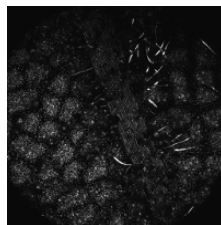


## Imaging Capabilities

- Single-Channel Confocal Systems
  - Reflected-Light System for Examining Surface Structures
  - GFP System for Epi-Fluorescence Imaging
- Complete with All Accessories Needed for Confocal Imaging
- Full Frame 4096 x 4096 Pixel Images (Uni-Directional Scan)
- 884  $\mu\text{m}$  x 884  $\mu\text{m}$  Field of View at 20X Magnification (FN25)
- 2 FPS for 512 x 512 Pixel Bi-Directional Scans
- Galvo-Galvo Scan Head with User-Selectable 1.0 to 10  $\mu\text{s}$  Dwell Times



Click for Full Size 2048 x 2048 Image  
Dragonfly Eye



Click for Full Size 1024 x 1024 Image  
Leaf Vein

400  $\mu\text{m}$  Deep Z-Projections of a Dragonfly Eye and Leaf Vein Imaged using the CM100 Reflected-Light Confocal Microscope. Both images were taken at a 1.3 mm x 1.3 mm field size.



Sam Tesfai  
General Manager,  
Thorlabs Imaging Systems

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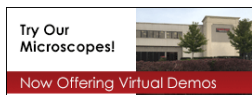
## Microscope Features

- Confocal Scan Path with Galvo-Galvo Scanners
- One Benchtop Excitation Laser
- One Multialkali Photomultiplier Tube (PMT)
- Filter Set for Reflected-Light or GFP Fluorescence Imaging
- One Objective and Matched  $\text{\O}75$   $\mu\text{m}$  Pinhole
- Upright Microscope Based on the Cerna<sup>®</sup> DIY Microscopy Platform Enables Upgrades to Add Functionality:
  - D1N Dovetail on Top of Scan Path Accepts an Epi-Illumination Module or a Cerna Widefield Imaging Module
  - 95 mm Dovetail on Rail Accepts Cerna Accessories
- Computer with National Instruments<sup>™</sup> (NI) PXIe-6363 X Series DAQ Card
- ThorImage<sup>®</sup>LS Data Acquisition Software with Lifetime Support

**Single-Channel Confocal Microscope Comparison<sup>a</sup>**

Item #	CM100	CM201
<b>Microscope Type</b>	Reflected Light	GFP Fluorescence
<b>Laser</b>	S1FC660 660 nm SM Laser (Red)	S4FC488 488 nm SM Laser (Blue)
<b>Scan Head</b>	Galvo-Galvo	
<b>Objective</b>	RMS20X 20X <sup>b</sup> Olympus Objective	N20X-PF 20X Nikon Objective
<b>Pinhole</b>	$\text{\O}75$ $\mu\text{m}$ , Optimized for Included Objective	
<b>PMT Detection</b>	One PMT1001/M Multialkali PMT	

- See the *Specs* tab for complete specifications.
- The RMS20X objective provides an effective magnification of 22.2X when used with the Confocal System as it is designed for a 180 mm tube lens and Thorlabs' microscopes use a 200 mm tube lens focal length. See the *Magnification & FOV* tab for more information on calculating effective objective magnification.



Thorlabs' Upgradeable Single-Channel Confocal Microscopes are complete upright confocal systems. By eliminating signals that originate from outside the focal plane, confocal microscopy provides the ability to acquire high-resolution, optically sectioned images from within a thick sample or to reduce background fluorescence from thin cultures. The CM100 supports confocal reflection imaging, which can be used for viewing the surface structure of biological samples or for inspection applications. The CM201 is optimized for imaging fluorescence produced by green fluorescent protein (GFP). Each system includes a laser, PMT detector, objective, and motorized Z-axis control. See the table to the right for a comparison of key features.

The top panel of each microscope has a female D1N dovetail that allows an epi-illumination module to be added to the system (see below), or Cerna widefield viewing accessories. The microscope body uses the same 95 mm dovetail as our Cerna system, making it easy to integrate trans-illumination modules, sample holders, or custom modules using body attachments built from our large catalog of optomechanics.

The CM100 reflected-light confocal microscope has a PFR14-P02 35 mm x 52 mm x 3 mm silver-coated mirror at the front of the scan path that directs light from the scanners to the objective. In order to use this microscope for widefield as well as confocal imaging, the user can replace this mirror with a beamsplitter or dichroic. The CM201 GFP confocal microscope features a movable silver-coated mirror on a manual slider at the front of the scan path that allows users to select

between confocal and widefield imaging modalities without replacing the optic. The body of each confocal microscope also includes a dual-objective changer, allowing the user to easily switch between the 20X objective included for confocal imaging and a user-provided second objective for widefield imaging.

Each microscope includes a PC with a National Instruments (NI) PXIe-6363 X Series DAQ card (see the *Specs* tab for details) and ThorImageLS data acquisition software. ThorImageLS was developed in conjunction with our laser scanning microscopy systems to provide a seamless, logical, intuitive program for acquiring and analyzing images. This open-source software package enables synchronization of external hardware and events, multi-dimensional data acquisition and display, region-of-interest scanning, and multi-user operation. All images are saved in the standard TIFF image format so that they can be viewed using software packages such as ImageJ/Fiji. See the *ThorImageLS* tab for additional information on ThorImageLS features. Upon the purchase of a confocal system, Thorlabs provides lifetime support for the ThorImageLS package.

These systems can be installed by the user and detailed instructions are provided in each manual (included with each system and accessible by clicking on the red documents icon below). An optional installation service is available for these systems. Use our Confocal Microscopy Contact Form for more information.

Specifications for Thorlabs' Upgradeable, Cerna®-Based Confocal Systems available from stock are provided here. If you are interested in a system with different specifications than those listed below, contact our sales team and applications engineers using our Confocal Microscopy Contact Form or at (703) 651-1700.

Item #	CM100	CM201
<b>System Type</b>	Reflected Light Imaging	GFP Fluorescence Imaging
<b>Excitation</b>		
<b>Item #</b>	S1FC660 Single Mode Fiber-Coupled Laser	S4FC488 Single Mode Fiber-Coupled Laser
<b>Wavelength</b>	660 nm	488 nm
<b>Max Output Power</b>	15 mW (Min)	16 mW (Min)
<b>Power Control</b>	Manual or 0 to 5 V External Signal	
<b>Scanning</b>		
<b>Scan Head</b>	Galvo-Galvo	
<b>Mirror</b>	PFR14-P02 35 mm x 52 mm x 3 mm Mirror, Protected Silver Coated with $\lambda/4$ Surface Flatness (Peak to Valley)	
<b>Digitization / Sampling Density</b>	Up to 4096 x 4096 Pixels (Uni-Directional Acquisition) Up to 2048 x 2048 Pixels (Bi-Directional Acquisition)	
<b>Scanning Speed</b>	Up to 2 FPS for 512 x 512 Pixel Bi-Directional Scans with 1 $\mu$ s Pixel Dwell Time	
<b>Pixel Dwell Time</b>	1.0 - 10.0 $\mu$ s, Software Selectable	
<b>Scan Zoom</b>	1X to 32X (Continuous)	
<b>Diffraction-Limited Field of View</b>	FN25: 796 $\mu$ m x 796 $\mu$ m @ 22.2X <sup>a</sup> 442 $\mu$ m x 442 $\mu$ m @ 40X <sup>b</sup> FN23: 733 $\mu$ m x 733 $\mu$ m @ 22.2X <sup>a</sup> 407 $\mu$ m x 407 $\mu$ m FOV @ 40X <sup>b</sup> (Field Number is Software Selectable up to FN25)	FN25: 884 $\mu$ m x 884 $\mu$ m @ 20X 442 $\mu$ m x 442 $\mu$ m @ 40X <sup>b</sup> FN23: 814 $\mu$ m x 814 $\mu$ m @ 20X 407 $\mu$ m x 407 $\mu$ m FOV @ 40X <sup>b</sup> (Field Number is Software Selectable up to FN25)
<b>Detection</b>		
<b>Pinhole</b>	$\varnothing$ 75 $\mu$ m, Optimized for Included 20X Objective	
<b>Photomultiplier Tube (PMT)</b>	PMT1001/M Multialkali PMT	
<b>Filters</b>	BSW10R 50:50 Beamsplitter WPQ10E-670 Quarter-Wave Plate LPVISE100-A Polarizers (2)	MD498 Dichroic: Refl. Band = 452 - 490 nm, Trans. Band = 505 - 800 nm MF525-39 Emission Filter: 525 nm / 39 nm
<b>Objective</b>		
<b>Item #</b>	RMS20X Olympus Plan Achromat Objective	N20X-PF Nikon Plan Fluorite Objective
<b>Magnification</b>	20X <sup>a</sup>	20X
<b>NA</b>	0.4	0.5
<b>Working Distance</b>	1.2 mm	2.1 mm
<b>Parfocal Length</b>	45.06 mm	60 mm
<b>Design Tube Lens Focal Length</b>	180 mm <sup>a</sup>	200 mm
<b>Coverslip Correction</b>	0.17 mm	
<b>Threading</b>	RMS	M25 x 0.75
<b>Fiber Patch Cables</b>		
<b>Laser to Scan Head</b>	P1-630PM-FC-2 2 m PM Patch Cable, 620 - 850 nm, FC/PC Connectors	P1-405B-FC-2 2 m SM Patch Cable, 405 - 532 nm, FC/PC Connectors
<b>Pinhole to Detector</b>	FG910UEC MM Fiber, 1 m, Armored Stainless Steel Protective Tubing, AR-Coated End Faces, SMA Connectors	
<b>General Microscope Features</b>		
<b>Widefield Viewing</b>	Silver-Coated Mirror can be Removed or Replaced with a Beamsplitter for Widefield Imaging	Silver-Coated Mirror on a Manual Slider to Switch Between Confocal and Widefield Imaging
	Female D1N Dovetail on Top of Scan Path to Mount Cerna Widefield Viewing Accessories	
<b>Microscope Body</b>	95 mm Dovetail Rail to Mount Cerna Body Attachments, Transmitted Illumination Modules, and Other Accessories 7.74" Throat Depth	
<b>Nosepiece</b>	CSN200 Dual Objective Changer ZFM2020 Focusing Module with 1" Fine Z Translation M32 x 0.75 Objective Threads (Two Places) M32 x 0.75 to M25 x 0.75 (Qty. 2) and M25 x 0.75 to RMS (Qty. 2) Adapters Included	
<b>Data Acquisition</b>		
<b>Type</b>	National Instruments PXIe-6363 X Series DAQ Card	

Item #	CM100	CM201
Analog Output <sup>c</sup>	4 Channels Resolution: 16 Bits Voltage Range: ±10 V Accuracy: 1.89 mV Update Rate: 2.86 MS/s	
Analog Input <sup>c</sup>	1 Channel Resolution: 16 Bits Voltage Range: ±10 V Accuracy: 1.66 mV	
Digital I/O <sup>c</sup>	48 Bidirectional Channels	
Clock Rate	10 MHz (Max)	
Frame In/Out Triggering	TTL	
Line Trigger Out	TTL	
Counter/Timers <sup>c</sup>	4	
<b>Computer and Software</b>		
Computer	PC with DAQ	
Software	ThorImage <sup>®</sup> LS with Lifetime Support	

- a. The RMS20X objective provides an effective magnification of 22.2X when used with the Confocal System as it is designed for a 180 mm tube lens and Thorlabs' microscopes use a 200 mm tube lens focal length. See the *Magnification & FOV* tab for more information on calculating effective objective magnification.
- b. For reference only. The included pinhole is not optimized for use with 40X objectives.
- c. All digital channels and one counter are available for use by the user. All other channels and counters are either actively used by the confocal system or reserved to support future upgrades.

Thorlabs recognizes that each imaging application has unique requirements.  
If you have any feedback, questions, or need a quotation, please contact  
ImagingSales@thorlabs.com or call (703) 651-1700.

**CM100 Reflected-Light Confocal Microscope**

Item # CM100 consists of:

- Single-Channel Reflected-Light Confocal Microscope Body
- Galvo-Galvo Scanner Control Box
- S1FC660 660 nm Single-Channel Fiber-Coupled Laser Source
- KST101 Stepper Motor Controller with USB Cable and KPS201 Power Supply
- PMT1001/M Multialkali PMT
- PH082E Post Holder, TR20/M Post, and CF175 Clamping Fork
- RMS20X 20X Olympus Plan Achromat Objective
- P1-630PM-FC-2 2 m Polarization Maintaining Patch Cable, FC/PC Connectors
- FG910UEC Multimode Fiber in Armored Patch Cable with Stainless Steel Protective Tubing, SMA Connectors
- 120" Long SMA to BNC Cable for Connecting PMT to NI Breakout Box
- Alignment Tool
- R1L3S3P Grid Slide
- NI Breakout Box with NI SHC68-68-EPM and NI SH6868 Cables
- Computer with 24" Monitor, Keyboard, and Mouse
- All Hardware Required for Mounting to an Optical Table
  - 12 1/4"-20 and 12 M6 Socket Head Cap Screws
  - Six 1/4" (M6) Washers
  - 1.5 mm, 2 mm, and 2.5 mm Hex Wrenches
  - 2 mm, 5 mm, 3/32" and 3/16" Hex L-Keys



Click to Enlarge  
CM100 Components  
Grid Slide, Alignment Tool, Computer Keyboard, and Mouse  
are Not Shown

**CM201 GFP Confocal Microscope**

Item # CM201 consists of:

- Single-Channel Fluorescence Confocal Microscope Body
- Galvo-Galvo Scanner Control Box
- S4FC488 488 nm Single-Channel Fiber-Coupled Laser Source
- KST101 Stepper Motor Controller with USB Cable and KPS201 Power Supply
- PMT1001/M Multialkali PMT
- PH082E Post Holder, TR20/M Post, and CF175 Clamping Fork
- N20X-PF 20X Nikon Plan Fluorite Objective
- P1-405B-FC-2 2 m Single Mode Patch Cable, FC/PC Connectors
- FG910UEC Multimode Fiber in Armored Patch Cable with Stainless Steel Protective Tubing, SMA Connectors
- 120" Long SMA to BNC Cable for Connecting PMT to NI Breakout Box
- Alignment Tool
- Fluorescent Bead Slide with 0.1, 0.2, 0.5, 1.0, and 4.0  $\mu\text{m}$  Bead Sizes
- Fluorescence Slide
- NI Breakout Box with NI SHC68-68-EPM and NI SH6868 Cables
- Computer with 24" Monitor, Keyboard, and Mouse
- All Hardware Required for Mounting to an Optical Table
  - 12 1/4"-20 and 12 M6 Socket Head Cap Screws
  - Six 1/4" (M6) Washers
  - 1.5 mm, 2 mm, and 2.5 mm Hex Wrenches
  - 2 mm, 5 mm, 3/32" and 3/16" Hex L-Keys



Click to Enlarge  
CM201 Components  
Fluorescent Bead Slide, Fluorescence Slide, Alignment Tool,  
Computer Keyboard, and Mouse are Not Shown

### Available Upgrades

Each Single-Channel Confocal Microscope is designed so that imaging capabilities can be added to accommodate new experimental needs as your research requirements grow. Some upgrades, such as adding Cerna® widefield imaging accessories, can be easily performed by the user. Others, such as adding additional excitation wavelengths, require replacing part of the hardware, in which case assistance will be provided by our technical staff. Contact us using the link in the green box to the right for additional information.

Four-Channel Upright Confocal Systems that incorporate the Confocal Capability Upgrades listed below are also available.

### Widefield Imaging Add-Ons

- Widefield Epi-Illumination with a Six-Filter Removable Turret or Using a Single Filter Cube
- Eyepieces, Trinoculars, and Scientific Cameras for Sample Viewing
- Transmitted Illumination Including Brightfield, Dodt Contrast, and DIC Imaging Modules
- Modules for Mounting Objective Changers, Condensers, and Sample Holders
- Microscope Mover with 2" of Translation in X and Y

### Confocal Capability Upgrades

- Up to 4 Excitation Wavelengths
- Up to 4 Confocal Detection Channels with Multialkali or GaAsP PMTs
- Change the Primary Mirror to a Dichroic or Movable Mirror (CM100 Only)
- Add a Motorized Shutter in the Scan Path
- Motorized Pinhole Wheel with 16 Round Sizes from  $\text{Ø}25\ \mu\text{m}$  to  $\text{Ø}2\ \text{mm}$
- Piezo Objective Stage or Z-Axis Piezo Sample Stage for Fast Z-Stacks
- Fast Motorized XY Stage for Large Area Tiling



[Click to Enlarge](#)

Single-Channel Confocal Microscope Upgraded with Epi-Fluorescence Imaging Functionality. The D1N dovetail on the top of the housing allows Cerna accessories to be added, such as the WFA2001 epi-illumination module, WFA4100 camera tube, and a scientific camera shown here. A silver-coated mirror mounted on a slider in the confocal scan head allows the user to switch between imaging modalities.



Sam Tesfai  
General Manager,  
Thorlabs Imaging Systems

**Questions?**

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## ThorImage<sup>®</sup>LS Software



ThorImageLS is an open-source image acquisition program that controls Thorlabs' microscopes, as well as supplementary external hardware. From prepared-slice multiphoton Z-stacks to simultaneous *in vivo* photoactivation and imaging, ThorImageLS provides an integrated, modular workspace tailored to the individual needs of the scientist. Its workflow-oriented interface supports single image, Z-stacks, time series, and image streaming acquisition, visualization, and analysis. See the video to the lower right for a real-time view of data acquisition and analysis with ThorImageLS.

ThorImageLS is included with a Thorlabs microscope purchase and open source, allowing full customization of software features and performance. ThorImageLS also includes Thorlabs' customer support and regular software updates to continually meet the imaging demands of the scientific community.

The full source code for ThorImage<sup>®</sup>LS is available for owners of a Bergamo<sup>®</sup>, Cerna<sup>®</sup>, Veneto<sup>®</sup> or confocal microscope. [Click here to receive your copy.](#)

For additional details, see the full web presentation.

### Advanced Software Functionality

- Multi-Column Customizable Workspace
- Image Acquisition Synced with Hardware Inputs and Timing Events
- Live Image Correction and ROI Analysis
- Independent Galvo-Galvo and Galvo-Resonant Scan Areas and Geometries
- Tiling for High-Resolution Large-Area Imaging
- Independent Primary and Secondary Z-Axis Control for Fast Deep-Tissue Scans
- Automated Image Capture with Scripts
  - Compatible with ImageJ Macros
- Multi-User Settings Saved for Shared Workstations
- Individual Colors for Detection Channels Enable Simple Visual Analysis

### Seamless Integration with Experiments

- Simultaneous Multi-Point Photoactivation and Imaging with Spatial Light Modulator
- Fast Z Volume Acquisition with PFM450E or Third-Party Objective Scanners
- Electrophysiology Signaling
- Wavelength Switching with Tiberius<sup>®</sup> Laser or Coherent Chameleon Lasers
- Pockels Cell ROI Masking
- Power Ramped with Depth to Minimize Damage and Maximize Signal-to-Noise

### New Functionality: Version 4.0 (Click to Expand for More Details)

Please contact [ImagingTechSupport@thorlabs.com](mailto:ImagingTechSupport@thorlabs.com) to obtain the latest ThorImageLS version compatible with your microscope. Because ThorImageLS 4.0 adds significant new features over 3.x, 2.x and 1.x versions, it may not be compatible with older microscopes. We continue to support older software versions for customers with older hardware. See the full web presentation for functionality of previous versions.

[Contact Us](#)

#### New Hardware Support

- Added Support for Windows<sup>®</sup> 10 OS
- Added Support for CS895MU and CS505MU Monochrome Cameras (Requires ThorCAM 3.2)
  - Allows for Hot Pixel Correction
- Added Support for CSN210 Motorized Dual-Objective Nosepiece
  - Allows for Improved Objective Setup and Control
- Added Support for Secondary Three Channel Controller
- Added Support for Second LED of the DC2200 LED Driver
- Added Support for New Version of Thorlabs' Tiberius<sup>®</sup> Femtosecond Ti:Sapphire Laser (Up to 1060 nm)
- Added Support for Second Channel for GGNI (Allows for Sequential Imaging with 2 Channels)
- Added Support for Controlling Up to 6 Digital Shutters (ThorShutterDig)
- Added Support for Resonant-Galvo-Galvo Scan-Head (Galvo-Resonant or Galvo-Galvo Scan Modes Only)
- Added Support for Coherent<sup>®</sup> Discovery with AOM Support (Requires Coherent<sup>®</sup> Discovery GUI Version 1.8.3 and 3rd Party Virtual Serial Port Software)

#### New Features

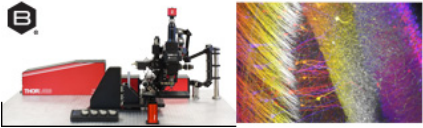


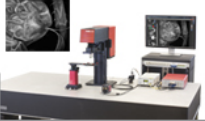

- Added Ability to Save Experiment Data in Multi-Page TIFF Format (OME TIFF)
- Added Rapid Image Update for Galvo-Galvo Scanner
  - Updates Image Every 16 Scan Lines During Acquisition
- Added Galvo-Galvo External Trigger Sync (Minimum 1 MHz) (GGNI Not Supported)
- Added Improved Galvo-Galvo and Galvo-Resonant Triggering Times
- Added Ability to Read Resonant Frequency Probe
- Added Configurable Trigger Output (Signal Generator) Based on Time or Other Digital Events
- Added Auto Update for Histograms
- Added Dedicated Bleach Shutter Control for Galvo-Galvo and GGNI
- Added Stimulation Epoch Control
- Added Additional Stimulation Features (Pre Idle, Post Idle) and Control Lines (Active, Cycle Output, Epoch)
- Added SLM Multiple Epoch Control (Random Epoch)
- Added Ability to Invert Z Control's Plus and Minus Buttons (Supports Both Primary and Secondary Z Controllers)
- Added Ability to Display X and Y Positions in Microns or Millimeters
- Added BCM-PA Slider Step Size

#### User Interface (UI) Improvements



- Renamed "Bleaching" to "Stimulation"
- Added Scale Bar in Image
- Added Help Menu Features
  - Allows User to Check for Updates
  - Allows User to View Log File for Troubleshooting
- Added Shortcuts to Hardware Settings and Application Settings in Hardware Connections Window and Edit Under Settings Menu
- Changed Capture Preview of Image to Show Averaged if Cumulative Mode is Used
- Added Control Digital Switches within Script
- Updated Digital Switch Configuration to be Saved in Experiment Settings and Viewable in Experiment Settings Browser
- Updated Extend Filing Numbering Index Out to 6 Digits
  - Allows for Setting Slide Step Size When Using Slider Plus and Minus Buttons for Power Adjustment.
- Added Auto Saving of Changed Fine Alignment Values
- Added Ability to Save Image Location and Zoom Level When Switching Image Modalities
- Added ThorSync Changes
  - Stack Panel Option
  - Virtual Channel

### Supported Imaging Platforms

 <p>Bergamo<sup>®</sup> II Multiphoton Microscopes</p>	 <p>DIY Multiphoton Microscope Kits</p>
 <p>Veneto<sup>®</sup> Inverted Microscopes</p>	 <p>Confocal Imaging Systems</p>
 <p>Hyperspectral Imaging System</p>	

## Laser Scanning Microscopy Tutorial

Laser scanning microscopy (LSM) is an indispensable imaging tool in the biological sciences. In this tutorial, we will be discussing confocal fluorescence imaging, multiphoton excitation fluorescence imaging, and second and third harmonic generation imaging techniques. We will limit our discussions to point scanning of biological samples with a focus on the technology behind the imaging tools offered by Thorlabs.

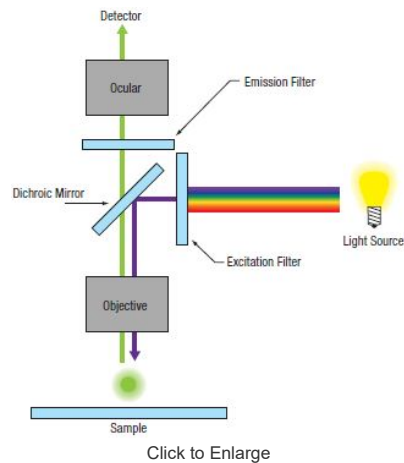
### Introduction

The goal of any microscope is to generate high-contrast, high-resolution images. In much the same way that a telescope allows scientists to discern the finest details of the universe, a microscope allows us to observe biological functioning at the nanometer scale. Modern laser scanning microscopes are capable of generating multidimensional data (X, Y, Z,  $\tau$ ,  $\lambda$ ), leading to a plethora of high-resolution imaging capabilities that further the understanding of underlying biological processes.

In conventional widefield microscopy (Figure 1, below left), high-quality images can only be obtained when using thin specimens (on the order of one to two cell layers thick). However, many applications require imaging of thick samples, where volume datasets or selection of data from within a specific focal plane is desired. Conventional widefield microscopes are unable to address these needs.

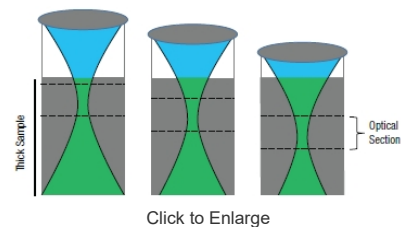
LSM, in particular confocal LSM and multiphoton LSM, allows for the visualization of thin planes from within a thick bulk sample, a technique known as optical sectioning. In confocal LSM, signals generated by the sample outside of the optical focus are physically blocked by an aperture, preventing their detection. Multiphoton LSM, as we will discuss later, does not generate any appreciable signal outside of the focal plane. By combining optical sectioning with incremented changes in focus (Figure 2, below right), laser scanning microscopy techniques can recreate 3D representations of thick specimen.

**Figure 1** Widefield Epi-Fluorescence

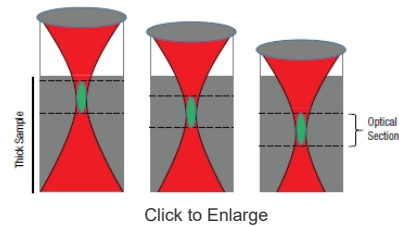


**Figure 2** Optical Sections (Visualization of Thin Planes within a Bulk Sample)

Optical Sectioning in Confocal Microscopy



### Optical Sectioning in Multiphoton Microscopy



Signal generated by the sample is shown in green. Optical sections are formed by discretely measuring the signal generated within a specific focal plane. In confocal LSM, out-of-focus light is rejected through the use of a pinhole aperture, thereby leading to higher resolution. In multiphoton LSM, signal is only generated in the focal volume. Signal collected at each optical section can be reconstructed to create a 3D image.

## Contrast Mechanisms in LSM

Biological samples typically do not have very good contrast, which leads to difficulty in observing the boundaries between adjacent structures. A common method for improving contrast in laser scanning microscopes is through the use of fluorescence.

In fluorescence, a light-emitting molecule is used to distinguish the constituent of interest from the background or neighboring structure. This molecule can already exist within the specimen (endogenous or auto-fluorescence), be applied externally and attached to the constituent (chemically or through antibody binding), or transfected (fluorescent proteins) into the cell.

In order for the molecule to emit light (fluoresce) it must first absorb light (a photon) with the appropriate amount of energy to promote the molecule from the ground state to the excited state, as seen in Figure 3A below. Light is emitted when the molecule returns back down to the ground state. The amount of fluorescence is proportional to the intensity ( $I$ ) of the incident laser, and so confocal LSM is often referred to as a linear imaging technique. Natural losses within this relaxation process require that the emitted photon have lower energy—that is, a longer wavelength—than the absorbed photon.

Multiphoton excitation (Figure 3B, below) of the molecule occurs when two (or more) photons, whose sum energy satisfies the transition energy, arrive simultaneously. Consequently, the two arriving photons will be of lower energy than the emitted fluorescence photon.

There are also multiphoton contrast mechanisms, such as harmonic generation and sum frequency generation, that use non-absorptive processes. Under conditions in which harmonic generation is allowed, the incident photons are simultaneously annihilated and a new photon of the summed energy is created, as illustrated in Figure 3C below.

Further constituent discrimination can be obtained by observing the physical order of the harmonic generation. In the case of second harmonic generation (SHG), signal is only generated in constituents that are highly ordered and lacking inversion symmetry. Third harmonic generation (THG) is observed at boundary interfaces where there is a refractive index change. Two-photon excitation and SHG are nonlinear processes and the signal generated is dependent on the square of the intensity ( $I^2$ ).

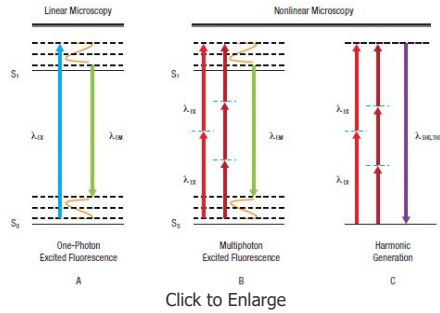
The nonlinear nature of signal generation in multiphoton microscopy means that high photon densities are required to observe SHG and THG. In order to accomplish this while maintaining relatively low average power on the sample, mode-locked femtosecond pulsed lasers, particularly Ti:Sapphire lasers, have become the standard.

Another consideration to be made in nonlinear microscopy is the excitation wavelength for a particular fluorophore. One might think that the ideal excitation wavelength is twice that of the one-photon absorption peak. However, for most fluorophores, the excited state selection rules are different for one- and two-photon absorption.

This leads to two-photon absorption spectra that are quite different from their one-photon counterparts. Two-photon absorption spectra are often significantly broader (can be  $>100$  nm) and do not follow smooth semi-Gaussian curves. The broad two-photon absorption spectrum of many fluorophores facilitates excitation of several fluorescent molecules with a single laser, allowing the observation of several constituents of interest simultaneously.

All of the fluorophores being excited do not have to have the same excitation peak, but should overlap each other and have a common excitation range. Multiple fluorophore excitation is typically accomplished by choosing a compromising wavelength that excites all fluorophores with acceptable levels of efficiency.

**Figure 3** Signal Generation in Laser Scanning Microscopy



#### Absorptive Process (A, B):

The absorption of one or more excitation photons ( $\lambda_{EX}$ ) promotes the molecule from the ground state ( $S_0$ ) to the excited state ( $S_1$ ). Fluorescence ( $\lambda_{EM}$ ) is emitted when the molecule returns to the ground state.

#### Non-Absorptive Process (C):

The excitation photons ( $\lambda_{EX}$ ) simultaneously convert into a single photon ( $\lambda_{SHG,THG}$ ) of the sum energy and half (for SHG) or one-third (for THG) the wavelength.



## Image Formation

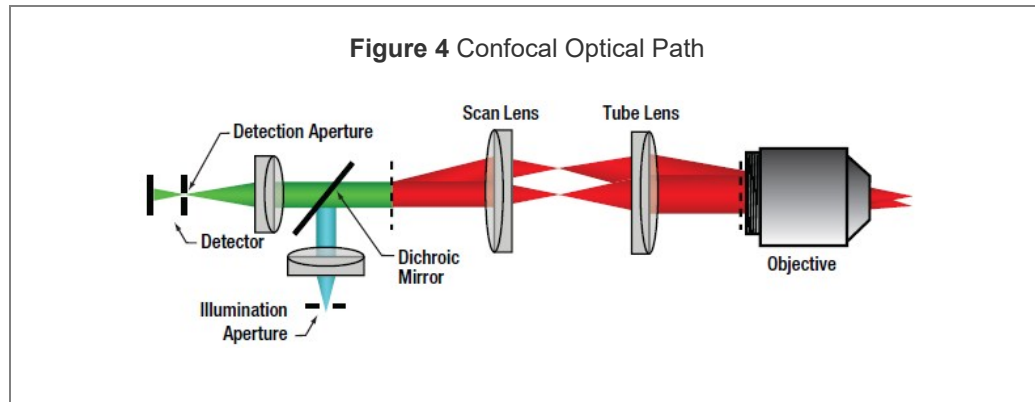
In a point-scanning LSM, the single-plane image is created by a point illumination source imaged to a diffraction-limited spot at the sample, which is then imaged to a point detector. Two-dimensional *en face* images are created by scanning the diffraction-limited spot across the specimen, point by point, to form a line, then line by line in a raster fashion.

The illuminated volume emits a signal which is imaged to a single-element detector. The most common single-element detector used is a photomultiplier tube (PMT), although in certain cases, avalanche photodiodes (APDs) can be used. CCD cameras are not typically used in point-scanning microscopes, though are the detector of choice in multifocal (i.e. spinning disk confocal) applications.

The signal from the detector is then passed to a computer which constructs a two-dimensional image as an array of intensities for each spot scanned across the sample. Because no true image is formed, LSM is referred to as a digital imaging technique. A clear advantage of single-point scanning and single-point detection is that the displayed image resolution, optical resolution, and scan field can be set to match a particular experimental requirement and are not predefined by the imaging optics of the system.

## Confocal LSM

In confocal LSM, point illumination, typically from a single mode, optical-fiber-coupled CW laser, is the critical feature that allows optical sectioning. The light emitted from the core of the single mode optical fiber is collimated and used as the illumination beam



for scanning. The scan system is then imaged to the back aperture of the objective lens which focuses the scanned beam to a diffraction-limited spot on the sample. The signal generated by the focused illumination beam is collected back through the objective and passed through the scan system.

After the scan system, the signal is separated from the illumination beam by a dichroic mirror and brought to a focus. The confocal pinhole is located at this focus. In this configuration, signals that are generated above or below the focal plane are blocked from passing through the pinhole, creating the optically sectioned image (Figure 2, above). The detector is placed after the confocal pinhole, as illustrated in Figure 4 to the right. It can be inferred that the size of the pinhole has direct consequences on the imaging capabilities (particularly, contrast, resolution and optical section thickness) of the confocal microscope.

The lateral resolution of a confocal microscope is determined by the ability of the system to create a diffraction-limited spot at the sample. Forming a diffraction-limited spot depends on the quality of the laser beam as well as that of the scan optics and objective lens.

The beam quality is typically ensured by using a single mode optical fiber to deliver the excitation laser light as a Gaussian point source, which is then collimated and focused into a diffraction-limited beam. In an aberration-free imaging system, obtained by using the highest quality optical elements, the size of this focus spot, assuming uniform illumination, is a function of excitation wavelength ( $\lambda_{EX}$ ) and numerical aperture (NA) of the objective lens, as seen in Equation 1.

$$\text{Spot Size} = \frac{1.22\lambda_{EX}}{NA}$$

Equation 1 Spot Size

In actuality, the beam isn't focused to a true point, but rather to a bullseye-like shape. The spot size is the distance between the zeros of the Airy disk (diameter across the middle of the first ring around the center of the bullseye) and is termed one Airy Unit (AU). This will become important again later when we discuss pinhole sizes.

The lateral resolution of the imaging system is defined as the minimum distance between two points for them to be observed as two distinct entities. In confocal (and multiphoton) LSM, it is common and experimentally convenient to define the lateral resolution according to the full width at half maximum (FWHM) of the individual points that are observed.

Using the FWHM definition, in confocal LSM, the lateral resolution ( $R_{\text{lateral,confocal}}$ ) is:

$$R_{\text{lateral,confocal}} = \frac{0.51\lambda_{EX}}{NA}$$

Equation 2 Lateral Resolution, Confocal LSM

and the axial resolution ( $R_{\text{axial,confocal}}$ ) is:

$$R_{\text{axial,confocal}} = \frac{0.88\lambda_{EX}}{n - \sqrt{n^2 - (NA)^2}}$$

Equation 3 Axial Resolution, Confocal LSM

where  $n$  is the refractive index of the immersion medium.

It is interesting to note that in a confocal microscope, the lateral resolution is solely determined by the excitation wavelength. This is in contrast to widefield microscopy, where lateral resolution is determined only by emission wavelength.

To determine the appropriate size of the confocal pinhole, we multiply the excitation spot size by the total magnification of the microscope:

$$D_{\text{pinhole}} = M_{\text{objective}} \times M_{\text{scan head}} \times \text{Spot Size}$$

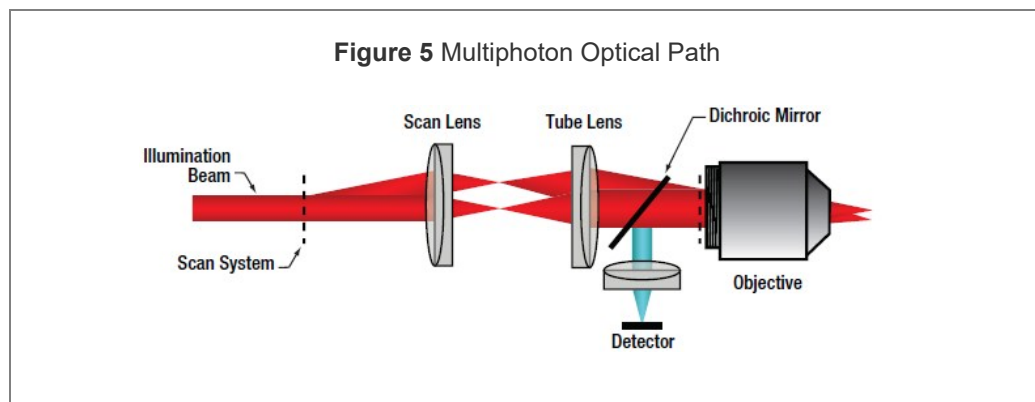
Equation 4 Pinhole Diameter

As an example, the appropriate size pinhole for a 60X objective with  $NA = 1.0$  for  $\lambda_{EX} = 488 \text{ nm}$  ( $M_{\text{scan head}} = 1.07$  for the Thorlabs Confocal Scan Head) would be  $38.2 \mu\text{m}$  and is termed a pinhole of 1 AU diameter. If we used the same objective parameters but changed the magnification to 40X, the appropriate pinhole size would be  $25.5 \mu\text{m}$  and would also be termed a pinhole of 1 AU diameter. Therefore, defining a pinhole diameter in terms of AU is a means of normalizing pinhole diameter, even though one would have to change the pinhole selection for the two different objectives.

Theoretically, the total resolution of a confocal microscope is a function of the excitation illumination spot size and the detection pinhole size. This means that the resolution of the optical system can be improved by reducing the size of the pinhole. Practically speaking, as we restrict the pinhole diameter, we improve resolution and confocality, but we also reduce the amount of signal reaching the detector. A pinhole of 1 AU is a good balance between signal strength, resolution, and confocality.

## Multiphoton LSM

In multiphoton LSM, a short pulsed free-space laser supplies the collimated illumination beam that passes through the scanning system and is focused by the objective. The very low probability of a multiphoton



absorption event

occurring, due to the  $I^2$  dependence of the signal on incident power, ensures signal is confined to the focal plane of the objective lens. Therefore, very little signal is generated from the regions above and below the focal plane. This effective elimination of out-of-focus signal provides inherent optical sectioning capabilities (Figure 2, above) without the need for a confocal pinhole. As a result of this configuration, the collected signal does not have to go back through the scanning system, allowing the detector to be placed as close to the objective as possible to maximize collection efficiency, as illustrated in Figure 5 to the right. A detector that collects signal before it travels back through the scan system is referred to as a non-descanned detector.

Again using the FWHM definition, in multiphoton LSM, the lateral resolution ( $R_{\text{lateral,multiphoton}}$ ) is:

$$R_{\text{lateral,multiphoton,NA}>0.7} = \frac{0.383\lambda_{EX}}{(NA)^{0.91}}$$

Equation 5 Lateral Resolution, Multiphoton LSM

and the axial resolution ( $R_{\text{axial,multiphoton}}$ ) is:

$$R_{\text{axial,multiphoton,NA}>0.7} = \frac{0.626\lambda_{EX}}{n - \sqrt{n^2 - (NA)^2}}$$

Equation 6 Axial Resolution, Multiphoton LSM

These equations assume an objective  $NA > 0.7$ , which is true of virtually all multiphoton objectives.

The longer wavelength used for multiphoton excitation would lead one to believe (from Equation 5) that the resolution in multiphoton LSM, compared to confocal LSM, would be reduced roughly by a factor of two. For an ideal point object (i.e. a sub-resolution size fluorescent bead) the  $I^2$  signal dependence reduces the effective focal volume, more than offsetting the 2X increase in the focused illumination spot size.

We should note that the lateral and axial resolutions display a dependence on intensity. As laser power is increased, there is a corresponding increase in the probability of signal being generated within the diffraction-limited focal volume. In practice, the lateral resolution in a multiphoton microscope is limited by how tightly the illumination beam can be focused and is well approximated by Equation 5 at moderate intensities. Axial resolution will continue to degrade as excitation power is increased.

## Image Display

Although we are not directly rendering an image, it is still important to consider the size of the image field, the number of pixels in which we are displaying our image (capture resolution) on the screen, and the lateral resolution of the imaging system. We use the lateral resolution because we are rendering an *en face* image. In order to faithfully display the finest features the optical system is capable of resolving, we must appropriately match resolution (capture and lateral) with the scan field. Our capture resolution must, therefore, appropriately sample the optical resolution.

In LSM, we typically rely on Nyquist sampling rules, which state that the pixel size should be the lateral resolution divided by 2.3. This means that if we take our 60X objective from earlier, the lateral resolution is 249 nm (Equation 2) and the pixel size in the displayed image should be 108 nm. Therefore, for a 1024 x 1024 pixel capture resolution, the scan field on the specimen would be ~111  $\mu\text{m}$  x 111  $\mu\text{m}$ . It should be noted that the 40X objective from our previous example would yield the exact same scan field (both objectives have the same NA) in the sample. The only difference between the two images is the angle at which we tilt our scanners to acquire the image.

It may not always be necessary to render images with such high resolution. We can always make the trade-off of image resolution, scan field, and capture resolution to create a balance of signal, sample longevity, and resolution in our images.

## Considerations in Live Cell Imaging

One of LSM's greatest attributes is its ability to image living cells and tissues. Unfortunately, some of the by-products of fluorescence can be cytotoxic. As such, there is a delicate balancing act between generating high-quality images and keeping cells alive.

One important consideration is fluorophore saturation. Saturation occurs when increasing the laser power does not provide the expected concurrent increase in the fluorescence signal. This can occur when as few as 10% of the fluorophores are in the excited state.

The reason behind saturation is the amount of time a fluorophore requires to relax back down to the ground state once excited. While the fluorescence pathways are relatively fast (hundreds of ps to a few ns), this represents only one relaxation mechanism. Triplet state conversion and nonradiative decay require significantly longer relaxation times. Furthermore, re-exciting a fluorophore before it has relaxed back down to the ground state can lead to irreversible bleaching of the fluorophore. Cells have their own intrinsic mechanisms for dealing with the cytotoxicity associated with fluorescence, so long as excitation occurs slowly.

One method to reduce photobleaching and the associated cytotoxicity is through fast scanning. While reducing the amount of time the laser spends on a single point in the image will proportionally decrease the amount of detected signal, it also reduces some of the bleaching mechanisms by allowing the fluorophore to completely relax back to the ground state before the laser is scanned back to that point. If the utmost in speed is not a critical issue, one can average several lines or complete frames and build up the signal lost from the shorter integration time.

The longer excitation wavelength and non-descanned detection ability of multiphoton LSM give the ability to image deeper within biological tissues. Longer wavelengths are less susceptible to scattering by the sample because of the inverse fourth power dependence ( $I^{-4}$ ) of scattering on wavelength. Typical penetration depths for multiphoton LSM are 250 - 500  $\mu\text{m}$ , although imaging as deep as 1 mm has been reported in the literature, compared to  $\sim 100 \mu\text{m}$  for confocal LSM.

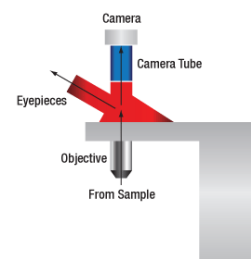
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## Magnification and Sample Area Calculations

### Magnification

The magnification of a system is the multiplicative product of the magnification of each optical element in the system. Optical elements that produce magnification include objectives, camera tubes, and trinocular eyepieces, as shown in the drawing to the right. It is important to note that the magnification quoted in these products' specifications is usually only valid when all optical elements are made by the same manufacturer. If this is not the case, then the magnification of the system can still be calculated, but an effective objective magnification should be calculated first, as described below.

Magnification and FOV Calculator



To adapt the examples shown here to your own microscope, please use our Magnification and FOV Calculator, which is available for download by clicking on the red button above. Note the calculator is an Excel spreadsheet that uses macros. In order to use the calculator, macros must be enabled. To enable macros, click the "Enable Content" button in the yellow message bar upon opening the file.

When viewing an image with a camera, the system magnification is the product of the objective and camera tube magnifications. When viewing an image with trinoculars, the system magnification is the product of the objective and eyepiece magnifications.

#### Example 1: Camera Magnification

When imaging a sample with a camera, the image is magnified by the objective and the camera tube. If using a 20X Nikon objective and a 0.75X Nikon camera tube, then the image at the camera has  $20X \times 0.75X = 15X$  magnification.

#### Example 2: Trinocular Magnification

When imaging a sample through trinoculars, the image is magnified by the objective and the eyepieces in the trinoculars. If using a 20X Nikon objective and Nikon trinoculars with 10X eyepieces, then the image at the eyepieces has  $20X \times 10X = 200X$  magnification. Note that the image at the eyepieces does not pass through the camera tube, as shown by the drawing to the right.

Manufacturer	Tube Lens Focal Length
Leica	f = 200 mm
Mitutoyo	f = 200 mm
Nikon	f = 200 mm
Olympus	f = 180 mm
Thorlabs	f = 200 mm
Zeiss	f = 165 mm

The rows highlighted in green denote manufacturers that do not use f = 200 mm tube lenses.

### Using an Objective with a Microscope from a Different Manufacturer

Magnification is not a fundamental value: it is a derived value, calculated by assuming a specific tube lens focal length. Each microscope manufacturer has adopted a different focal length for their tube lens, as shown by the table to the right. Hence, when combining optical elements from different manufacturers, it is necessary to calculate an *effective* magnification for the objective, which is then used to calculate the magnification of the system.

The effective magnification of an objective is given by Equation 1:

$$\text{Effective Objective Magnification} = \text{Design Magnification} \times \frac{f_{\text{Tube Lens in Microscope}} \text{ (mm)}}{f_{\text{Design Tube Lens of Objective}} \text{ (mm)}} \quad (\text{Eq. 1})$$

Here, the Design Magnification is the magnification printed on the objective,  $f_{\text{Tube Lens in Microscope}}$  is the focal length of the tube lens in the microscope you are using, and  $f_{\text{Design Tube Lens of Objective}}$  is the tube lens focal length that the objective manufacturer used to calculate the Design Magnification. These focal lengths are given by the table to the right.

Note that Leica, Mitutoyo, Nikon, and Thorlabs use the same tube lens focal length; if combining elements from any of these manufacturers, no conversion is needed. Once the effective objective magnification is calculated, the magnification of the system can be calculated as before.

#### Example 3: Trinocular Magnification (Different Manufacturers)

When imaging a sample through trinoculars, the image is magnified by the objective and the eyepieces in the trinoculars. This example will use a 20X Olympus objective and Nikon trinoculars with 10X eyepieces.

Following Equation 1 and the table to the right, we calculate the effective magnification of an Olympus objective in a Nikon microscope:

$$\text{Effective Objective Magnification} = 20X \times \frac{200 \text{ mm}}{180 \text{ mm}} = 22.2X$$

The effective magnification of the Olympus objective is 22.2X and the trinoculars have 10X eyepieces, so the image at the eyepieces has  $22.2X \times 10X = 222X$  magnification.

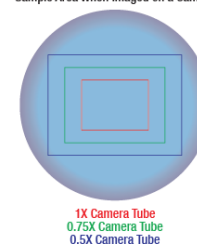
### Sample Area When Imaged on a Camera

When imaging a sample with a camera, the dimensions of the sample area are determined by the dimensions of the camera sensor and the system magnification, as shown by Equation 2.

$$\text{Sample Area (mm} \times \text{mm)} = \frac{\text{Camera Sensor Width (mm)}}{\text{System Magnification}} \times \frac{\text{Camera Sensor Height (mm)}}{\text{System Magnification}} \quad (\text{Eq. 2})$$

The camera sensor dimensions can be obtained from the manufacturer, while the system magnification is the multiplicative product of the objective magnification and the camera tube magnification (see Example 1). If needed, the objective magnification can be adjusted as shown in Example 3.

Sample Area When Imaged on a Camera





As the magnification increases, the resolution improves, but the field of view also decreases. The dependence of the field of view on magnification is shown in the schematic to the right.

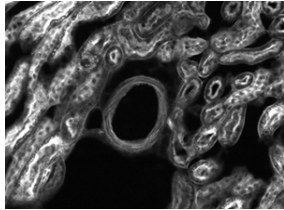
*Example 4: Sample Area*

The dimensions of the camera sensor in Thorlabs' previous-generation 1501M-USB Scientific Camera are 8.98 mm × 6.71 mm. If this camera is used with the Nikon objective and trinoculars from Example 1, which have a system magnification of 15X, then the image area is:

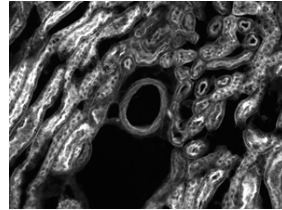
$$\text{Sample Area (mm} \times \text{mm)} = \frac{8.98 \text{ mm}}{15X} \times \frac{6.71 \text{ mm}}{15X} = 599 \mu\text{m} \times 447 \mu\text{m}$$

**Sample Area Examples**

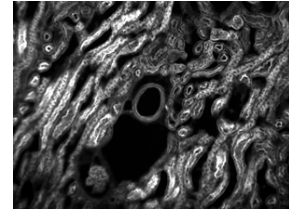
The images of a mouse kidney below were all acquired using the same objective and the same camera. However, the camera tubes used were different. Read from left to right, they demonstrate that decreasing the camera tube magnification enlarges the field of view at the expense of the size of the details in the image.



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Acquired with 1X Camera Tube (Item # WFA4100)



[Click to Enlarge](#)  
Acquired with 0.75X Camera Tube (Item # WFA4101)



[Click to Enlarge](#)  
Acquired with 0.5X Camera Tube (Item # WFA4102)

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**Selected Confocal Microscopy Publications****2020**

Caccavano, A. et al. Inhibitory Parvalbumin Basket Cell Activity is Selectively Reduced during Hippocampal Sharp Wave Ripples in a Mouse Model of Familial Alzheimer's Disease. *The Journal of Neuroscience* 5116–5136 (2020).

**2016**

Lewin, A. E. et al. Optogenetic and pharmacological evidence that somatostatin-GABA neurons are important regulators of parasympathetic outflow to the stomach. *The Journal of Physiology* 2661–2679 (2016). doi:10.1113/JP272069

**2015**

Dechen, K., Richards, C. D., Lye, J. C., Hwang, J. E. C. & Burke, R. Compartmentalized zinc deficiency and toxicities caused by ZnT and Zip gene over expression result in specific phenotypes in *Drosophila*. *The International Journal of Biochemistry & Cell Biology* 60, 23–33 (2015).

Jia, Y. et al. Activation of platelet protease-activated receptor-1 induces epithelial-mesenchymal transition and chemotaxis of colon cancer cell line SW620. *Oncol. Rep.* 33, 2681–2688 (2015).

Lu, W. et al. Inhibiting the mobilization of Ly6Chigh monocytes after acute myocardial infarction enhances the efficiency of mesenchymal stromal cell transplantation and curbs myocardial remodeling. *Am J Transl Res* 7, 587–597 (2015).

Lu, W. et al. Photoluminescent Mesoporous Silicon Nanoparticles with siCCR2 Improve the Effects of Mesenchymal Stromal Cell Transplantation after Acute Myocardial Infarction. *Theranostics* 5, 1068–1082 (2015).

Zuo, S., Hughes, M. & Yang, G.-Z. Novel Balloon Surface Scanning Device for Intraoperative Breast Endomicroscopy. *Ann Biomed Eng* 1–14 (2015). doi:10.1007/s10439-015-1493-2

**2014**

Brown, C. M., Melcher, J. T. & Stranick, S. J. Scan Linearization for Resonant Optomechanical Systems. in IM1C.3 (OSA, 2014). doi:10.1364/ISA.2014.IM1C.3

Partridge, J. G., Lewin, A. E., Yasko, J. R. & Vicini, S. Contrasting actions of group I metabotropic glutamate receptors in distinct mouse striatal neurones. *J Physiol* 592, 2721–2733 (2014).

Qin, X., Qiu, C. & Zhao, L. Maslinic acid protects vascular smooth muscle cells from oxidative stress through Akt/Nrf2/HO-1 pathway. *Mol Cell Biochem* 390, 61–67 (2014).

9.Liu, J. et al. Oleanolic Acid Suppresses Aerobic Glycolysis in Cancer Cells by Switching Pyruvate Kinase Type M Isoforms. *PLOS ONE* 9, e91606 (2014).

**2013**

Hao, X. et al. Contrast reversal confocal microscopy. *Optics Communications* 298–299, 272–275 (2013).

Lalchandani, R. R., Goes, M.-S. van der, Partridge, J. G. & Vicini, S. Dopamine D2 Receptors Regulate Collateral Inhibition between Striatal Medium Spiny Neurons. *J. Neurosci.* 33, 14075–14086 (2013).

## Standard Mechanical Interfaces on DIY Cerna® Components

The table below gives the dovetail, optical component threads, and cage system interfaces that are present on each DIY Cerna component. If a DIY Cerna component does not have one of the standard interfaces in the table, it is not listed here. Please note that mechanical compatibility does not ensure optical compatibility. Information on optical compatibility is available from Thorlabs' web presentations.

Item #	Microscope Dovetails										Optical Component Threads <sup>a</sup>		Cage Systems <sup>b</sup>
	95 mm	D1N	D2N	D2NB	D3N	D5N	D1T	D3T	D1Y	D5Y	Internal	External	
2CM1	-	-	-	-	-	-	-	-	-	-	SM1 <sup>c</sup> (1.035"-40) and SM2 <sup>d</sup> (2.035"-40)	SM1 <sup>c</sup> (1.035"-40)	60 mm <sup>d</sup>
2CM2	-	-	-	-	-	-	-	-	-	-	SM1 <sup>c</sup> (1.035"-40) and SM2 <sup>d</sup> (2.035"-40)	SM1 <sup>c</sup> (1.035"-40)	30 mm <sup>c</sup>
BSA2000 <sup>e</sup>	-	-	-	-	Female	-	-	-	-	-	-	-	-
CEA1350	Male	Female	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CEA1400	Male	Female	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CEA1500	Male	Female	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CEA1600	Male	Female	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CFB1500	Male	-	-	-	-	-	-	-	-	-	-	-	-
CSA1000	Female	-	-	-	-	-	-	-	-	-	-	-	-
CSA1001	Female	-	-	-	-	-	-	-	-	-	SM1 <sup>c</sup> (1.035"-40)	-	30 mm <sup>c</sup>
CSA1002	Female	-	-	-	-	-	-	-	-	-	SM2 <sup>d</sup> (2.035"-40)	-	60 mm <sup>d</sup>
CSA1003	-	Female	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CSA1051	Female	-	-	-	-	-	-	-	Male	-	-	-	-
CSA1200 <sup>e,f</sup>	-	-	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CSA1400 <sup>e</sup>	-	-	-	-	-	-	Female	-	-	-	-	-	60 mm <sup>d</sup>
CSA1500 <sup>e,g</sup>	-	-	-	-	-	-	-	-	-	-	-	-	-
CSA2000 <sup>e</sup>	-	-	-	-	Female	-	-	-	-	-	SM2 <sup>d</sup> (2.035"-40)	-	60 mm <sup>d</sup>
CSA2001	-	-	-	-	Female	-	-	-	-	-	-	SM2 <sup>d</sup> (2.035"-40)	-
CSA2100 <sup>e</sup>	-	-	-	-	-	-	-	-	-	-	SM2 <sup>d</sup> (2.035"-40)	-	60 mm <sup>d</sup>
CSA3000(/M)	-	Male	-	-	-	-	-	-	-	-	-	-	-
CSA3010(/M)	-	Male	-	-	-	-	-	-	-	-	-	-	30 mm <sup>c</sup> and 60 mm <sup>d</sup>
Item #	95 mm	D1N	D2N	D2NB	D3N	D5N	D1T	D3T	D1Y	D5Y	Internal	External	Cage Systems
CSC1001	-	-	-	-	Male	-	-	-	-	-	-	-	-
CSC1002	-	-	-	-	Male	-	-	-	-	-	-	-	-
CSC2001	-	-	-	-	Male	-	-	-	-	-	-	-	-
CSD1001	-	Male & Female	-	Female	-	-	-	-	-	-	-	-	-
CSD1002	-	Male & Female	-	-	-	-	-	-	-	-	-	C-Mount <sup>h</sup>	-
CSE2000	-	Male & Female	-	-	-	-	-	-	-	-	-	-	60 mm <sup>d</sup>
CSE2100	-	Male & Female	-	-	-	-	-	Female	-	-	SM1 <sup>c</sup> (1.035"-40)	-	30 mm <sup>c</sup> and 60 mm <sup>d</sup>
CSE2200	-	Male & Female	-	-	-	-	-	Female	-	-	SM1 <sup>c</sup>	-	30 mm <sup>c</sup> and



XT95P12/(M)	Female	-	-	-	-	-	-	-	-	-	-	-	-	-
ZFM1020	Female	-	-	-	-	-	-	-	-	-	-	-	-	-
ZFM1030	Female	-	-	-	-	-	-	-	-	-	-	-	-	-
ZFM2020	Female	-	-	-	-	-	-	-	-	-	-	-	-	-
ZFM2030	Female	-	-	-	-	-	-	-	-	-	-	-	-	-

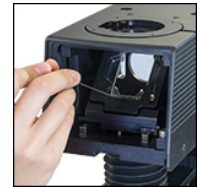
- Thorlabs' optical component thread adapters can be used to convert between C-Mount threads, SM1 threads, SM2 threads, and virtually every other optical thread standard.
- Our cage system size adapters and drop-in adapter can be used to convert between 16 mm, 30 mm, and 60 mm cage systems.
- Our 30 mm cage plates can convert between SM1 lens tubes and 30 mm cage systems.
- Our 60 mm cage plates can convert between SM2 lens tubes and 60 mm cage systems.
- Attach to a ZFM focusing module to add a female 95 mm dovetail.
- The CSA1200 mounting arm is compatible with the CSN1201 and CSN1202 nosepieces.
- This blank arm is designed for custom DIY machining for non-standard components, threads, and bores.
- C-Mount and CS-Mount standards feature the same 1.00"-32 threads, but C-Mounts have a 5 mm longer flange-to-sensor distance.

### Reflected-Light Imaging Confocal Microscope

- ▶ Ideal for Imaging Surface Structures of Biological Samples and for Inspection Applications
- ▶ Red 660 nm Single Mode Laser
- ▶ Silver-Coated Mirror at Front of Scan Path can be Removed or Replaced with a Dichroic to Enable Widefield Imaging
- ▶ Compatible with DIY Cerna Platform for Later Upgrades
- ▶ Optical Table, Sample Holder, and Rack Sold Separately



Click to Enlarge



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The front panel of the microscope can be removed to access the mirror at the front of the scan path. The user may replace this mirror with a beamsplitter or dichroic, enabling widefield imaging with the addition of DIY Cerna components.

Thorlabs' CM100 Reflected-Light Confocal Microscope includes a 660 nm single mode laser, galvo-galvo scanner, a removable silver-coated mirror at the front of the scan path, polarizers and a quarter-wave plate to minimize unwanted reflections for improved image quality, and a single multi-alkali PMT. A manual dual objective changer with motorized focus control accepts the included 20X objective. Each system includes a computer with DAQ and the ThorImage<sup>®</sup>LS data acquisition software.

This microscope includes everything needed for reflected-light confocal imaging. Additionally, the 95 mm dovetail on the microscope body and female D1N dovetail on top of the scan path allow the user to upgrade the microscope with widefield imaging capabilities using our wide selection of DIY Cerna components. The included dual objective changer allows the user to easily switch between the included 20X objective and a second user-provided objective with a different magnification when moving between widefield and confocal imaging.

The microscope body features the same 7.74" throat depth as our widefield Cerna systems. To achieve this large free space beneath and around the objective, the microscope is not designed to be free-standing and must be bolted to an optical table. For this purpose, 1/4"-20 and M6 cap screws are included to provide compatibility with both metric and imperial optical tables.



Due to a major supply chain outage, we are currently unable to confirm a shipment date for Item # CM100. We anticipate having more information to share in late 2022.

Part Number	Description	Price	Availability
CM100	Single-Channel Confocal Microscope for Reflected-Light Imaging	\$40,737.72	Lead Time

## GFP Fluorescence Confocal Microscope

- ▶ Ideal for Green Fluorescent Protein (GFP) Microscopy
- ▶ Blue 488 nm Single Mode Laser
- ▶ Silver-Coated Mirror on Manual Slider at Front of Scan Path for Switching Between Imaging Modalities
- ▶ Compatible with DIY Cerna Platform for Later Upgrades
- ▶ Optical Table, Sample Holder, and Rack Sold Separately



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The D1N dovetail on the top of the housing allows Cerna accessories to be added, such as the WFA2001 epillumination module, WFA4100 camera tube, and a scientific camera shown here. A silver-coated mirror mounted on a slider in the confocal scan head allows the user to switch between imaging modalities.

Thorlabs' CM201 GFP Fluorescence Confocal Microscope includes a 488 nm single mode laser, galvo-galvo scanner, a silver-coated mirror on a two position slider at the front of the scan path, a dichroic and emission filter for GFP confocal fluorescence imaging, and a single multi-alkali PMT. A manual dual objective changer with motorized focus control accepts the included 20X objective. Each system includes a computer with DAQ and the ThorImage<sup>®</sup>LS data acquisition software.

This microscope includes everything needed for GFP confocal imaging. Additionally, the 95 mm dovetail on the microscope body and female D1N dovetail on top of the scan path allow the user to upgrade the microscope with widefield imaging capabilities using our wide selection of DIY Cerna components. The included dual objective changer allows the user to easily switch between the included 20X objective and a second user-provided objective with a different magnification when moving between widefield and confocal imaging.

The microscope body features the same 7.74" throat depth as our widefield Cerna systems. To achieve this large free space beneath and around the objective, the microscope is not designed to be free-standing and must be bolted to an optical table. For this purpose, 1/4"-20 and M6 cap screws are included to provide compatibility with both metric and imperial optical tables.



Due to a major supply chain outage, we are currently unable to confirm a shipment date for Item # CM201. We anticipate having more information to share in late 2022.

Part Number	Description	Price	Availability
CM201	Single-Channel Confocal Microscope for GFP Fluorescence Imaging	\$47,645.57	Lead Time



## Epi-Illuminator Module with Removable Turret for Six Filter Sets



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The CSE2000 Epi-Illuminator Module features 4-40 taps to connect to a 60 mm cage system.

- ▶ Included Removable Turret Holds Up to Six Filter Sets without the Need for Filter Cubes
- ▶ Monitor Filter Turret Position on a PC Using Included Software Package
- ▶ Stackable Design with Female and Male D1N Dovetails on Top and Bottom, Respectively
- ▶ Directly Compatible with WFA3110 DIC Analyzer
- ▶ CSE2000 Do-It-Yourself Epi-Illuminator Module:
- ▶ Build Your Own Optical Path with 60 mm Cage Components
- ▶ CSE2200 Ready-to-Use Epi-Illuminator Module:
- ▶ Accepts a Collimated Illumination Source via a 60 mm Cage System or D3T Dovetail
- ▶ Additional Optical Path Available for 1" Lens Tubes or a 30 mm Cage System

### Turret Position Sensor Software

Version 4.0 (October 11, 2019)

This software package contains the installation files for the GUI interface, driver, SDK, and support documentation. The software is compatible with Windows® 7 (64 bit) and Windows 10 (64 bit) systems.



Click for Details

The CSE2200 provides a field stop diaphragm and conditioning optics for the main optical path as well as a side port for an additional optical path.

These Epi-Illuminator Modules expand the functionality of our Confocal Microscopes to include fluorescence and brightfield microscopy. With a female D1N dovetail on top and a male D1N dovetail on the bottom, they can be easily added to the microscopes above and accept Cerna widefield viewing modules. Up to six fluorescence filter sets may be installed directly into the turret, allowing for quick and simple switching between imaging different fluorophores without the need for filter cubes; the turret is also available for purchase separately. A mini-USB port on the side of the housing and included GUI software package allow the turret position to be monitored using a PC (not included); see the Software box at the top right for details. Each module also includes a manual shutter to block the illumination beam while not in use and a slot for the WFA3110 DIC analyzer for DIC microscopy.

### CSE2200 Ready-To-Use Epi-Illuminator Module

The CSE2200 was designed for use with microscopes that utilize a longer working distance over the epi-illumination path than standard Cerna microscopes, including the confocal microscopes available above. The collimating optics in the primary optical path condition incoming light to provide even widefield illumination at the sample plane. For epi-illumination modules compatible with our standard Cerna microscopes, see the Cerna Epi-Illumination web presentation.

The primary epi-illumination path features AR-coated optical elements designed for homogeneous illumination over 365 - 700 nm, as well as a field stop diaphragm. A collimated illumination source, such as our Solis® High-Power LED, can be attached using the female D3T dovetail on the back of the module and an SM2A56 adapter, available for purchase below. Light sources with a Ø3 mm or Ø5 mm liquid light guide (LLG) can also be utilized with a LLG collimation adapter. Alternatively, the dovetail can be removed, exposing 4-40 taps that allow a 60 mm cage system to be attached.

An additional optical path can also be utilized by inserting a beamsplitter or dichroic (30 mm x 42 mm, 2.0 ± 0.1 mm in thickness) into the module; please contact Tech Support for compatible dichroics or beamsplitters. The side port features internal SM1 (1.035"-40) threading as well as 4-40 taps for our 30 mm cage system.

### CSE2000 Epi-Illuminator Module for Home Builders

The CSE2000 does not include beam conditioning optics, allowing home builders to incorporate custom illumination sources in a Cerna microscope. This standalone module features four 4-40 taps for 60 mm cage systems on the back side. This mechanical interface enables Thorlabs' extensive selection of 60 mm cage components to be used to build custom epi-illumination paths. To utilize the CSE2000 with an existing microscope featuring alternate dovetails, contact Tech Support for a custom solution.

Specifications		Less [-]	
Item #	CSE2000	CSE2200	
Module Type	DIY	Ready to Use	
Fluorescence Filter Set Mounting	Turret for Six Filter Sets (Included)		
Dovetails	Male D1N (Bottom) Female D1N (Top)	Male D1N (Bottom) Female D1N (Top) Female D3T (Back)	
Compatible Illumination Source	User-Constructed Optical Path <sup>a</sup>	Primary Path: Collimated Side Port: User-Constructed	
Cage System Compatibility	60 mm Cage System (4-40 Tap, 4 Places)	Primary Path: 60 mm Cage System (4-40 Tap, 4 Places) <sup>b</sup> Side Port: 30 mm Cage System (4-40 Tap, 4 Places)	
Distance from the Secondary Image Plane to the Side Port (D1)	-	122 mm (See <a href="#">Optical Diagram</a> )	
Distance from the Male D1N Dovetail to the Objective Back Focal Plane (D2)	-	150 mm (See <a href="#">Optical Diagram</a> )	
Control Voltage Connector	Mini USB Port		
Input Voltage	5 V DC		
Field Stop Diaphragm	-	Ø0.8 mm - Ø14.0 mm	
Accepted Optics (Not Included)	Filter Turret	Ø25.4", <5.1 mm Thick Emission/Excitation Filters 25 mm x 36 mm, 1 ± 0.1 mm Thick Dichroics/Beamsplitters	
	Dichroic/Beamsplitter Holder	-	30 mm x 42 mm, 2.0 ± 0.1 mm Thick Optic
Weight	1.7 kg	4.4 kg	
Operating Temperature	5 to 35 °C		
Storage Temperature	-20 °C to +50 °C		
Dimensions (L x W x H)	146.3 mm x 132.7 mm x 90.6 mm (5.76" x 5.22" x 3.57")	478.0 mm x 132.7 mm x 91.1 mm (18.82" x 5.22" x 3.59")	

a. The CSE2000 does not come with a preconfigured optical path to accept an illumination source. It is designed for users who want to design their own optical path.

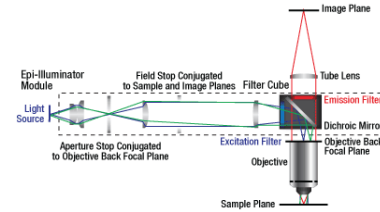
b. Accessible by Removing D3T Dovetail Back Piece

Part Number	Description	Price	Availability
CSE2200	Epi-Illuminator Module with Turret for Six Filter Sets, Conditioning Optics, Additional Optical Port, WD = 150 mm	\$6,195.00	Lead Time
CSE2000	Epi-Illuminator Module with Turret for Six Filter Sets	\$3,485.74	Today
SM2A56	Adapter with Male D3T Dovetail and External SM2 Threads	\$114.18	Today

## Epi-Illuminator Modules for One Filter Cube

- ▶ WFA2001: Pre-Installed Optical Path that Accepts an Uncollimated Illumination Source with a Ø3 mm Core Liquid Light Guide or SM1 Threads
- ▶ WFA2002: Build Your Own Optical Path using SM1 and 30 mm Cage Components
- ▶ Magnetic Door Cover Holds One Olympus-Compatible Filter Cube
- ▶ Stackable Design with Female and Male D1N Dovetails on Top and Bottom, Respectively
- ▶ MDFM-MF2 Filter Cube is Sold Separately Below

Installation of the MDFM-MF2 Filter Cube (Sold Below) with a Filter Set into the WFA2001 Single-Cube Epi-Illuminator Module



Click for Details  
Optical Diagram for WFA2001 (See *Optical Diagrams* Tab for More Details)

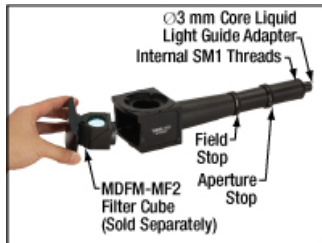
These epi-illumination modules hold one filter cube in the epi-illumination pathway of a DIY Cerna system. With a female D1N dovetail on top and a male D1N dovetail on the bottom, they are designed to mate with the epi-illumination arm of a Cerna microscope body, as well as with each other. Additional details on dovetails are available in the *Microscope Dovetails* tab.

The WFA2001 Epi-Illuminator Module's optical input port has internal SM1 (1.035"-40) threads. For this port, we include two adapters: an adapter that accepts a Ø3 mm core liquid light guide and an SM1T10 adapter that accepts an uncollimated light source with an internally SM1-threaded output, such as a mounted LED. For use with a Ø5 mm core liquid light guide, the AD5LLG adapter is available for purchase separately. This module is designed so that evenness of illumination is optimized at a 110 mm working distance between the bottom of the dovetail and the back focal plane of the objective; see the optical diagram to the right. Its pre-installed optical path consists of collimating optics that are AR coated for 350 - 700 nm, as well as field stop and aperture stop diaphragms (Item # SM1D12D) that can be adjusted from Ø0.8 mm to Ø12.0 mm (Ø0.03" to Ø0.47").

In contrast, the WFA2002 Epi-Illuminator Module does not include the pre-installed optical path of the WFA2001. As shown by the photo below, removing the optical path exposes internal SM1 threads and four 4-40 taps for 30 mm cage systems on the back of the module. These mechanical interfaces enable Thorlabs' extensive selection of SM1 and 30 mm cage components to be used to build custom epi-illumination paths.

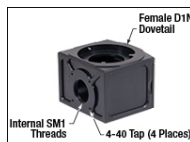
Each of these modules includes a magnetically secured cover that can be connected to an MDFM-MF2 Filter Cube (sold separately below), as shown by the animation in the upper right. Extra covers, which the user can attach to the filter cubes to speed up filter cube exchange, are sold as Item # WFA2001C.

Please note that these epi-illumination modules do not have the slot required for the WFA3110 DIC Analyzer, so a custom mounting solution will be needed. The CSE2100, CSE2000, and CSE2200 Epi-Illuminator Modules sold below are directly compatible with this DIC analyzer.



Click to Enlarge

The WFA2001 Epi-Illuminator Module has a ready-to-use beam path, with field stop and aperture stop diaphragms, for a mounted LED or another uncollimated source with SM1 threads or a Ø3 mm liquid light guide.



Click to Enlarge



Click to Enlarge

The back of the WFA2002 Epi-Illuminator Module has internal SM1 threads and four 4-40 taps for a 30 mm cage system, which can be used to support home-built epi-illumination setups.

Part Number	Description	Price	Availability
WFA2001	Epi-Illuminator Module for 1 Cube, Conditioning Optics, Male & Female D1N Dovetails	\$1,978.70	Today
WFA2002	Epi-Illuminator Module for 1 Cube, Male & Female D1N Dovetails	\$451.02	Today
WFA2001C	Extra Filter Cube Cover for WFA2001 and WFA2002 Epi-Illuminator Modules	\$202.64	Today

## Turret for Six Filter Sets

- ▶ Removable Turret Compatible with CSE2000, CSE2100, and CSE2200 Epi-Illuminator Modules
- ▶ Easily Install Multiple Filter Sets in a Particular Configuration
- ▶ Designed for Use with Thorlabs' and Third-Party Fluorescence Filter Sets

The CSE2000W Turret allows for simple installation of up to six fluorescence filter sets, without the need for filter cubes. This turret directly accepts up to six filter sets without requiring filter cubes: six excitation filters (Ø25.4 mm, <5.1 mm in thickness), six emission filters (Ø25.4 mm, <5.1 mm in thickness), and six rectangular optics (25 mm x 36 mm, 1 ± 0.1 mm in thickness). The emission filters are mounted at a 5° angle to reduce unwanted back reflections. Having multiple filter sets in the same turret, or multiple turrets with particular optic configurations, makes it easy to switch amongst fluorescence filter sets, beamsplitters with crossed polarizers for reflected light imaging, and mirrors. When used with the CSE2000, CSE2100, or CSE2200 epi-illumination modules (sold above), the turret position can be monitored remotely on a PC with software included with those modules.

The circular optic apertures feature internal SM1 (1.035"-40) threading for simple mounting of Ø1" optical elements; each turret ships with twelve SM1RR retaining rings, one to secure each circular filter. To install the rectangular optics, remove the top plate of the turret by loosening the three M3 screws, then remove the leaf springs to secure each optic. The turret utilizes grip holes on either side of each filter set, both for ease of use and to ensure the optical elements are not touched once installed. Once inserted into an epi-illumination module, simply turn the exposed knurled wheel to switch the filter set in the optical path.



Click to Enlarge  
CSE2000W Filter Turret with Top Plate

Click for Details  
Remove the Top Plate from the Turret to Install Dichroics or Mirrors

Part Number	Description	Price	Availability
CSE2000W	Turret for Six Filter Sets	\$2,337.42	Today

## OEM Microscopy Filter Cube

- ▶ OEM Filter Cube from Olympus
- ▶ Designed for Use with Thorlabs' and Third-Party Fluorescence Filter Sets

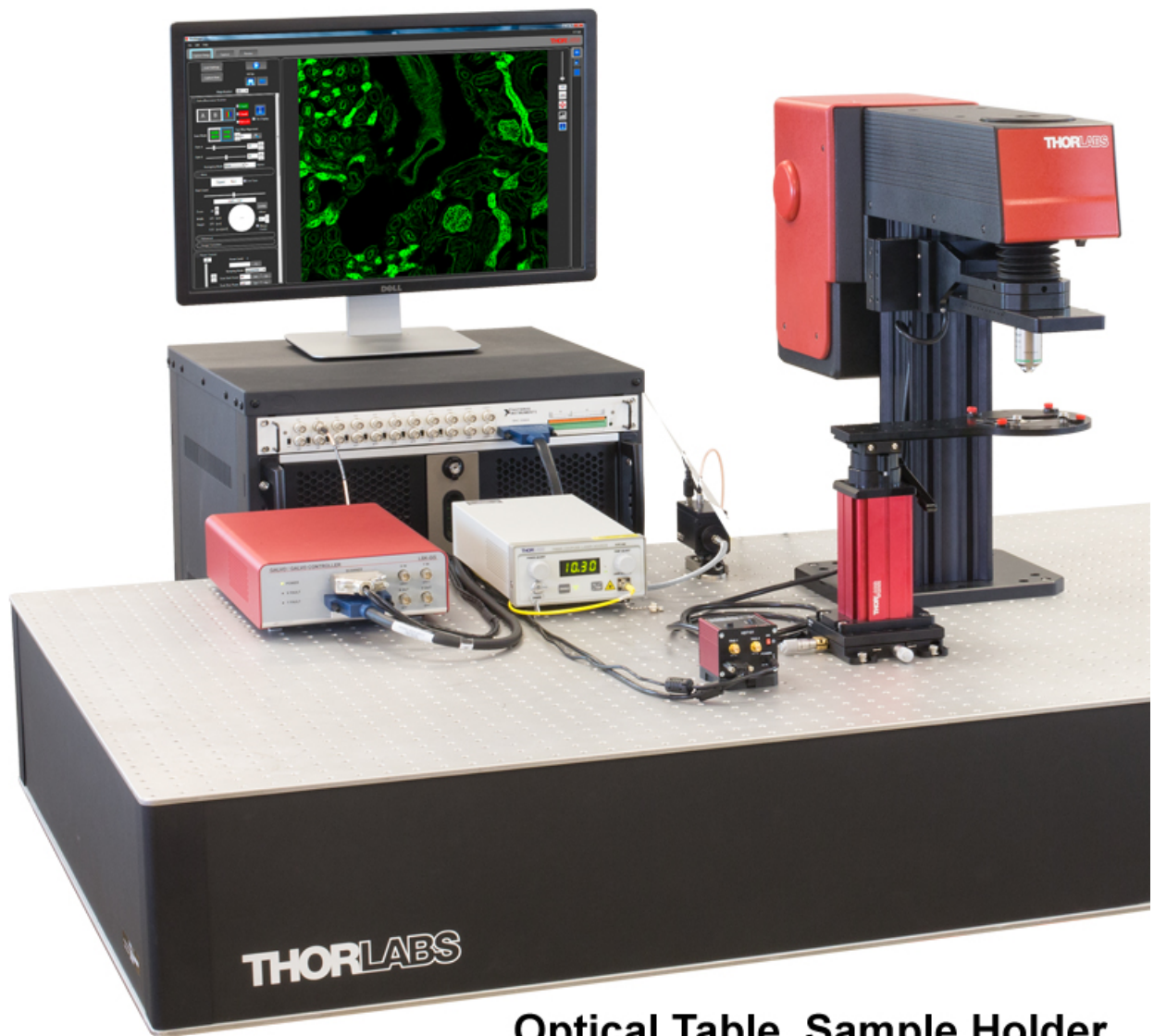
This OEM Filter Cube manufactured by Olympus can be used with the WFA2001. The filter cube holds one set of fluorescence filters: an excitation filter (Ø25 mm, up to 5 mm thick), an emission filter (Ø25 mm, up to 3.5 mm thick), and a dichroic mirror (up to 25.2 mm x 36.0 mm x 1.0 mm). Optics can be mounted, aligned, and swapped out easily as illustrated in the video to the right. The filter cube body may be disassembled for optic installation or removal using a Phillips head screwdriver. For detailed assembly instructions, please refer to the assembly manual in the table below.

This filter cube is also available with pre-installed filter sets.

Item #	Manufacturer Part #	Microscope Manufacturer	Compatible Microscopes	Assembly Manual
MDFM-MF2	Olympus U-MF2	Olympus	AX, BX2, and IX2 series	MDFM-MF2 Manual
		Thorlabs	Cerna Epi-Fluorescence Microscopes with WFA2001 or WFA2002 Epi-Illuminator Module	

Please contact Tech Support with questions regarding other cube compatibility, mounting, and filter options. Thorlabs does not guarantee compatibility with other industry-standard microscopes not mentioned on this webpage.

Part Number	Description	Price	Availability
MDFM-MF2	OEM Microscopy Cube Assembly for Olympus AX, BX2, IX2, and Thorlabs Cerna Microscopes with WFA2001 or WFA2002 Epi-Illuminator Module	\$563.67	Today



**Optical Table, Sample Holder,  
and Rack Not Included**